

Product Data Sheet

Nuciferine

Cat. No.: HY-N0049

CAS No.: 475-83-2Molecular Formula: $C_{19}H_{21}NO_2$ Molecular Weight: 295.38

Target: 5-HT Receptor; Dopamine Receptor; Parasite

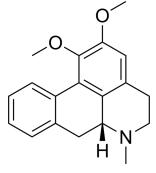
Pathway: GPCR/G Protein; Neuronal Signaling; Anti-infection

Storage: Powder -20°C 3 years

4°C 2 years

In solvent -80°C 2 years

-20°C 1 year



SOLVENT & SOLUBILITY

In Vitro

DMSO: 5 mg/mL (16.93 mM; ultrasonic and warming and heat to 60°C)

Preparing Stock Solutions	Solvent Mass Concentration	1 mg	5 mg	10 mg
	1 mM	3.3855 mL	16.9273 mL	33.8547 mL
	5 mM	0.6771 mL	3.3855 mL	6.7709 mL
	10 mM	0.3385 mL	1.6927 mL	3.3855 mL

Please refer to the solubility information to select the appropriate solvent.

In Vivo

- 1. Add each solvent one by one: 10% DMSO >> 40% PEG300 >> 5% Tween-80 >> 45% saline Solubility: ≥ 1.11 mg/mL (3.76 mM); Clear solution
- 2. Add each solvent one by one: 10% DMSO >> 90% corn oil Solubility: ≥ 1.11 mg/mL (3.76 mM); Clear solution

BIOLOGICAL ACTIVITY

	Description	Nuciferine is an antagonist at 5-HT $_{2A}$ (IC $_{50}$ =478 nM), 5-HT $_{2C}$ (IC $_{50}$ =131 nM), and 5-HT $_{2B}$ (IC $_{50}$ =1 μ M), an inverse agonist at 5-HT $_{1A}$ (IC $_{50}$ =150 nM), a partial agonist at D $_{2}$ (EC $_{50}$ =64 nM), D $_{5}$ (EC $_{50}$ =2.6 μ M) and 5-HT $_{6}$ (EC $_{50}$ =700 nM), an agonist at 5-HT $_{1A}$ (EC $_{50}$ =3.2 μ M) and D $_{4}$ (EC $_{50}$ =2 μ M) receptor.			
IC	IC ₅₀ & Target	5-HT _{2C} Receptor 131 nM (IC ₅₀)	Schistosome	5-HT ₇ Receptor 150 nM (IC ₅₀)	5-HT _{2A} Receptor 478 nM (IC ₅₀)
		5-HT _{2B} Receptor 1 μM (IC ₅₀)	5-HT ₆ Receptor 700 nM (EC50)	5-HT _{1A} Receptor 3.2 μM (EC50)	D ₂ Receptor 64 nM (EC50)

	·	D_5 Receptor 2.6 μM (EC50)
In Vitro	Nuciferine is a partial agonist at DD_2 receptor with an activity (E_{max} =67% of dopamine) similar to aripiprazole (E_{max} =50% dopamine). In line with its partial agonist activity, Nuciferine inhibited dopamine-induced activation of G_i with a potency similar to clozapine (Nuciferine K_B =62 nM; Clozapine K_B =20 nM) as determined via Schild regression analysis ^[1] . The natural product Nuciferine acts as an effective inhibitor of adult worm motility. Nuciferine is effective at inhibiting both basal and HT evoked motility of adult schistosomes. Nuciferine inhibits Sm.5HTR _L and schistosomule with 0.24±0.04 and 0.62±0.22 prespectively ^[2] . MCE has not independently confirmed the accuracy of these methods. They are for reference only.	
In Vivo	effects of a 5-HT _{2A} agonist, substited activity, inhibited phencyclidine (inhibition without induction of casubstitution to PCP alone. In the cat 10 mg/kg Nuciferine (80.63% d lower doses tested (0.1 mg/kg-3 r	psychotic drug action, Nuciferine blocks head-twitch responses and discriminative stimulus ituted for clozapine discriminative stimulus, enhanced amphetamine induced locomotor (PCP)-induced locomotor activity, and rescued PCP-induced disruption of prepulse atalepsy. In the presence of 1 or 3 mg/kg Nuciferine, cumulative PCP doses produce similar clozapine-trained animals, a dose-dependent substitution for 1.25 mg/kg clozapine is seen rug lever responding), with an ED ₅₀ value of 5.42 mg/kg (95% CI 3.09-9.48 mg/kg) while the mg/kg) fails to produce substitution for clozapine's discriminative cue. In addition to a high clozapine-appropriate lever, 10 mg/kg Nuciferine also produces significant rate cle control points (p<0.001) ^[1] .

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PROTOCOL

Kinase Assay [1]

For affinity determination, Nuciferine is subjected to primary radioligand binding assays tested at a single $10~\mu M$ concentration to displace 50% of the radioligand at a given receptor target. If a more than 50% of the radioligand is displaced, Nuciferine is selected for a secondary binding assay tested at 11 concentrations in triplicate in competition with the radioligand to generate an IC_{50} and K_i . Binding assays are performed in 96-well plates with $125~\mu L$ per well in appropriate binding buffer using radioligand at or near the K_d . Plates are incubated at room temperature in the dark for 90 min. Reactions are stopped by vacuum filtrations onto 0.3% polyethyleneimine soaked 96-well filter mats using a 96-well Filtermate harvester, followed by at least three washes of cold wash buffer. Scintillation cocktail is melted onto dried filters and radioactivity is counted using a Wallac Trilux Microbeta^[1].

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Cell Assay [1]

Cells are plated into 48-well plates one day before uptake is performed. Cells are washed with 0.5 mL uptake buffer (4 mM Tris, 6.25 mM HEPES, 120 mM NaCl, 5 mM KCl, 1.2 mM CaCl $_2$, 1.2 mM MgSO $_4$, 5.6 mM D-glucose, 1.7 mM ascorbic acid, and 1 μ M pargyline, pH 7.4). Cells are incubated with 225 μ L uptake buffer with or without the indicated concentration of Nuciferine for 15 minutes. After incubation, 25 μ L uptake buffer containing 3 H-DA and DA is added for a final concentration of 20 nM 3 H-DA and 1 μ M DA. Cells are incubated at 37°C for 20 minutes or for the time indicated. Nonspecific uptake is determined in the presence of 10 μ M nomifensine. Uptake is terminated by aspirating uptake buffer and washing each well twice with 0.5 mL ice-cold uptake buffer. Cells are lysed in 0.1 N NaOH and transferred to vials containing 3 mL scintillation cocktail. Radioactivity is quantitated using a Beckman LS6500 counter. Data are analyzed in Graph Pad Prism 5.0 $^{[1]}$. MCE has not independently confirmed the accuracy of these methods. They are for reference only.

Animal Administration [1]

Mice^[1]

Adult male NIH Swiss mice weighing approximately 25 g are used. Mice are injected with either Nuciferine (1, 3, or 10 mg/kg, i.p.) or vehicle, n=4 mice/condition. Fifteen minutes later, mice are injected with 1 mg/kg DOI (i.p.) and immediately placed in an observation chamber (new cage without bedding). Head-twitches (operationally defined as a rapid rotational jerk of the head that can be distinguished from species-appropriate grooming or scratching behaviors) are counted for 20 minutes in 5 minute bins. For the time-course study, mice are pretreated with 3.0 mg/kg Nuciferine (i.p.) at 60, 45, 30, 15, or 0 minutes (co-injection) prior to the 1.0 mg/kg DOI (i.p.) injection, and head-twitches are counted as described above. In one experiment, mice (n=4 per condition) are pretreated with an injection (s.c.) of 3.0 mg/kg Nuciferine or vehicle 15 minutes

prior to 1.0 mg/kg DOI injection (i.p.) and head-twitches are counted as described above. All experiments are performed by 3 observers, with 2 observers blinded to the experimental conditions which are evenly distributed. Power analyses are performed with the resulting data. The two highest doses of Nuciferine tested (10 and 3 mg/kg), had 0.96 and 0.88 power to detect significance (α =0.05). As these experiments are performed blinded and in distinct mice, further replication is not performed.

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CUSTOMER VALIDATION

- Int Immunopharmacol. 2023 Apr 29;119:110204.
- J Funct Foods. August 2022, 105182.
- Nutr Metab. 2021 Feb 18;18(1):20.
- Mol Pharmacol. 2023 Nov;104(5):230-238.

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REFERENCES

[1]. Farrell MS, et al. In Vitro and In Vivo Characterization of the Alkaloid Nuciferine. PLoS One. 2016 Mar 10;11(3):e0150602.

[2]. Chan JD, et al. Pharmacological profiling an abundantly expressed schistosome serotonergic GPCR identifies nuciferine as a potent antagonist. Int J Parasitol Drugs Drug Resist. 2016 Dec;6(3):364-370.

Caution: Product has not been fully validated for medical applications. For research use only.

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