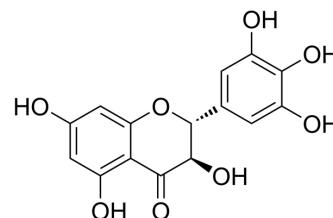


Dihydromyricetin

Cat. No.:	HY-N0112		
CAS No.:	27200-12-0		
Molecular Formula:	C ₁₅ H ₁₂ O ₈		
Molecular Weight:	320.25		
Target:	mTOR; Autophagy; Influenza Virus; DNA/RNA Synthesis		
Pathway:	PI3K/Akt/mTOR; Autophagy; Anti-infection; Cell Cycle/DNA Damage		
Storage:	Powder	-20°C	3 years
		4°C	2 years
	In solvent	-80°C	1 year
		-20°C	6 months



SOLVENT & SOLUBILITY

In Vitro

DMSO : ≥ 100 mg/mL (312.26 mM)
 * "≥" means soluble, but saturation unknown.

Preparing Stock Solutions	Solvent Concentration	Mass		
		1 mg	5 mg	10 mg
	1 mM	3.1226 mL	15.6128 mL	31.2256 mL
	5 mM	0.6245 mL	3.1226 mL	6.2451 mL
	10 mM	0.3123 mL	1.5613 mL	3.1226 mL

Please refer to the solubility information to select the appropriate solvent.

In Vivo

- Add each solvent one by one: 10% DMSO >> 40% PEG300 >> 5% Tween-80 >> 45% saline
 Solubility: ≥ 2.5 mg/mL (7.81 mM); Clear solution
- Add each solvent one by one: 10% DMSO >> 90% (20% SBE-β-CD in saline)
 Solubility: ≥ 2.5 mg/mL (7.81 mM); Clear solution
- Add each solvent one by one: 10% DMSO >> 90% corn oil
 Solubility: ≥ 2.5 mg/mL (7.81 mM); Clear solution

BIOLOGICAL ACTIVITY

Description

Dihydromyricetin is a potent inhibitor with an IC₅₀ of 48 μM on dihydropyrimidinase. Dihydromyricetin can activate autophagy through inhibiting mTOR signaling. Dihydromyricetin suppresses the formation of mTOR complexes (mTORC1/2). Dihydromyricetin is also a potent influenza RNA-dependent RNA polymerase inhibitor with an IC₅₀ of 22 μM.

IC₅₀ & Target

Dihydropyrimidinase 48 μM (IC ₅₀)	mTORC1	mTORC2	Autophagy
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In Vitro	<p>Dihydromyricetin, a flavonol, significantly inhibits the catalytic activities of dihydropyrimidinase toward both the natural substrate dihydrouracil and xenobiotic substrate 5-propyl-hydantoin. Dihydromyricetin exhibits a significant inhibitory effect on the activities of dihydropyrimidinase for both substrates, even more than Myricetin does. The IC_{50} values of Dihydromyricetin for dihydropyrimidinase determined from the titration curves using Dihydrouracil and 5-propyl-hydantoin are 48 ± 2 and 40 ± 2 μM, respectively^[1].</p> <p>Dihydromyricetin (DHM) supplementation significantly reverses the increased phosphorylation of mTOR at Ser²⁴⁴⁸ (p-mTOR) during D-gal administration, which suggests that Dihydromyricetin can activate autophagy through inhibiting mTOR signaling^[2].</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>
In Vivo	<p>Changes in learning and memory capacity in rats administrated normal control group, D-gal group, D-gal+Dihydromyricetin (100 mg/kg) group, D-gal+Dihydromyricetin (200 mg/kg) group assessed by morris water maze (MWM) (n=10 per group). Dihydromyricetin (DHM) treatment significantly shortens the escape latency when compared with D-gal-induced model group^[2].</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>

PROTOCOL

Kinase Assay ^[1]	<p>A rapid spectrophotometric assay is used to determine the enzymatic activity for hydantoinase, allantoinase, dihydroorotase, and imidase. Dihydrouracil, 5-propyl-hydantoin, and phthalimide are used as substrates. Unless explicitly stated otherwise, Dihydrouracil (2 mM) is used as the substrate in the standard assay of dihydropyrimidinase. Briefly, the decrease in absorbancy at 230, 248, and 298 nm is measured upon hydrolysis of Dihydrouracil, 5-propyl-hydantoin, and Phthalimide as the substrate at 25°C, respectively. To start the reaction, the purified dihydropyrimidinase (10-70 μg) is added to a 2 mL solution containing the substrate and 100 mM Tris-HCl (pH 8.0). Substrate hydrolysis is monitored with a UV/vis spectrophotometer. The extinction coefficient of each substrate is determined experimentally by direct measurement with a spectrophotometer. The extinction coefficients of Dihydrouracil, 5-propyl-hydantoin, and Phthalimide are $0.683 \text{ mM}^{-1}\text{cm}^{-1}$ at 230 nm, $0.0538 \text{ mM}^{-1}\text{cm}^{-1}$ at 248 nm, and $3.12 \text{ mM}^{-1}\text{cm}^{-1}$ at 298 nm, respectively. The initial rates of change are a function of enzyme concentration within the absorbance range of 0.01-0.18 min^{-1}. A unit of activity is defined as the amount of enzyme catalyzing the hydrolysis of 1 μmol substrate/min, and the specific activity is expressed in terms of units of activity per milligram of enzyme. The kinetic parameters K_m and V_{max} are determined from a non-linear plot by fitting the hydrolyzing rate from individual experiments to the Michaelis-Menten equation^[1].</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>
Cell Assay ^[2]	<p>Hippocampus and cortex tissue samples are homogenized in lysis buffer containing 20 mM Tris (pH 7.5), 135 mM NaCl, 2 mM EDTA, 2 mM DTT, 25 mM β-glycerophosphate, 2 mM sodium pyrophosphate, 10% glycerol, 1% Triton X-100, 1 mM sodium orthovanadate, 10 mM NaF, 10 μg/mL aprotinin, 10 μg/mL leupeptin, and 1 mM PMSF for 30 min on ice and centrifuged at $12000 \times g$ at 4°C for 30 min. The supernatant is collected and protein quantification is carried out using a BCA kit. The protein samples are boiled in the presence of sample buffer at 95°C for 5 min. The target protein is separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE), transferred to nitrocellulose membrane, and then probed by corresponding primary and secondary antibodies. Finally, the target protein is visualized by enhanced chemiluminescence (ECL) reagent exposure to X-ray film^[2].</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>
Animal Administration ^[2]	<p>Rats^[2]</p> <p>Totally 40 male Sprague-Dawley (SD) rats (age: 8 weeks old; body weight: 160 ± 20 g) are used. The rats are randomly divided into four groups including normal control group, D-gal model group, and D-gal combined with DHM at the doses of 100 and 200 mg/kg-d groups with 10 rats in each group. All rats are housed at the environment with room temperature of $22 \pm 2^\circ\text{C}$ and a dark-light cycle (12 h: 12h), and provided the accessibility to food and water ad libitum. After adapting to new environment for 1 week, the rats from DHM groups are administered with DHM dissolved in distilled water at the designated dosages by gavage once a day at 8:00am for 6 consecutive weeks. The rats from the normal control group are administrated with distilled water. Except from the normal control group, the rats from other groups are subjected to subcutaneous injection of D-gal at the dose of 150 mg/kg.d for 6 consecutive weeks. Each administration of DHM should be 2 h ahead of D-</p>

gal injection.

MCE has not independently confirmed the accuracy of these methods. They are for reference only.

CUSTOMER VALIDATION

- Acta Pharm Sin B. 2021 Jan;11(1):143-155.
- Mol Ther. 2019 May 8;27(5):1051-1065.
- Phytomedicine. 2023 Jul 26;119:154997.
- Phytomedicine. 2023 Mar 12.
- Phytomedicine. 2022 Mar 2;99:154027.

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REFERENCES

- [1]. Huang CY. Inhibition of a Putative Dihydropyrimidinase from *Pseudomonas aeruginosa* PAO1 by Flavonoids and Substrates of Cyclic Amidohydrolases. *PLoS One*. 2015 May 19;10(5):e0127634.
- [2]. Kou X, et al. Ampelopsin attenuates brain aging of D-gal-induced rats through miR-34a-mediated SIRT1/mTORsignal pathway. *Oncotarget*. 2016 Nov 15;7(46):74484-74495.
- [3]. Chang H, et al. Ampelopsin suppresses breast carcinogenesis by inhibiting the mTOR signalling pathway. *Carcinogenesis*. 2014 Aug;35(8):1847-54.
- [4]. Václav Zima, et al. Unraveling the Anti-Influenza Effect of Flavonoids: Experimental Validation of Luteolin and its Congeners as Potent Influenza Endonuclease Inhibitors. *Eur J Med Chem*. 22 August 2020, 112754.
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