Diphenyl Blue

HY-D0970	
72-57-1	
$C_{34}H_{24}N_6Na_4O_{14}S_4$	NAD, SP, SP, ONE NH ₂ OH, NH ₂ OH, NH ₂ OH, NH ₂
960.81	
Fluorescent Dye	
Others	NaO´´`o ó´`ONa
4°C, sealed storage, away from moisture and light * In solvent : -80°C, 6 months; -20°C, 1 month (sealed storage, away from moisture and light)	
	HY-D0970 72-57-1 C ₃₄ H ₂₄ N ₆ Na ₄ O ₁₄ S ₄ 960.81 Fluorescent Dye Others 4°C, sealed storage, away from moisture and light * In solvent : -80°C, 6 months; -20°C, 1 month (sealed storage, away from moisture and light)

SOLVENT & SOLUBILITY

DMSO: 25 mg/mL (26.02 mM; Need ultrasonic) In Vitro H₂O: 1 mg/mL (1.04 mM; ultrasonic and warming and heat to 60°C) Mass Solvent 5 mg 10 mg 1 mg Concentration Preparing 1 mM 1.0408 mL 5.2039 mL 10.4079 mL **Stock Solutions** 1.0408 mL 5 mM 0.2082 mL 2.0816 mL 10 mM 0.1041 mL 0.5204 mL 1.0408 mL Please refer to the solubility information to select the appropriate solvent.

BIOLOGICAL ACTIVITY Description Diphenyl Blue (Trypan Blue) is a cell active dye, the most commonly used dye for the identification of dead cells, of en used to test cell membrane integrity and cell viability. Diphenyl Blue staining is one of the methods for tissue and cell culture. When cells are deactivated or have incomplete cell membranes, Diphenyl Blue can stain them Blue. Normal living cells with intact cell membranes reject Diphenyl blue and do not stain them blue. However, macrophages are capable of phagocytosis of Diphenyl Blue, so it can be used as a living stain for macrophages^[1]. In Vitro Preparation of Trypan Blue working solution 1.1 Preparation of the stock solution Dissolve 10 mg of Trypan Blue in 100 mL 0.85% Nacl to obtain 0.4% Trypan Blue. Note: It is recommended to store the stock solution at -2022 -8022 away from light and avoid repetitive freeze-thaw cycles. 1.2 Preparation of Trypan Blue working solution Dilute the stock solution in 0.85% Nacl to obtain 0.04% of Trypan Blue working solution. Note: Please adjust the concentration of Trypan Blue working solution according to the actual situation. Cell staining 2.1 For suspension cells, centrifuge at 1000 g at 40 for 3-5 minutes and then discard the supernatant. Wash twice with PBS, 5

minutes each time.

For adherent cells, discard the cell culture medium, and add trypsin to dissociate cells to make a single-cell suspension. Centrifuge at 1000 g at 4 for 3-5 minutes and then discard the supernatant. Wash twice with PBS, 5 minutes each time.

 $2.2\,\text{Add}\,1\,\text{mL}$ of rypan Blue working solution, and then incubate at room temperature for 5 minutes.

2.3 Centrifuge at 400 g at 4 \boxtimes for 3-4 minutes and then discard the supernatant.

2.4 Wash twice with PBS, 5 minutes each time.

 ${\tt 2.5 \, Resuspend \, cells \, with \, serum-free \, cell \, culture \, medium \, or \, {\tt PBS}, \, and \, then \, detect \, by \, fluorescence \, microscope.}$

MCE has not independently confirmed the accuracy of these methods. They are for reference only.

CUSTOMER VALIDATION

- Cell Death Dis. 2023 Aug 29;14(8):573.
- Photochem Photobiol. 2023 May 22.

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REFERENCES

[1]. Daly, M. L., DeRosa, C. A., Kerr, C., Morris, W. A., & Fraser, C. L. (2016). Blue thermally activated delayed fluorescence from a biphenyl difluoroboron β-diketonate. RSC Advances, 6(85), 81631–81635. doi:10.1039/c6ra18374c

Caution: Product has not been fully validated for medical applications. For research use only.