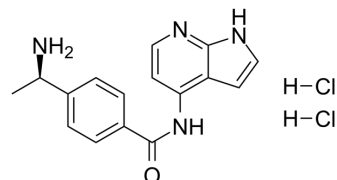


## Y-33075 dihydrochloride

<b>Cat. No.:</b>	HY-10069
<b>CAS No.:</b>	173897-44-4
<b>Molecular Formula:</b>	C <sub>16</sub> H <sub>18</sub> Cl <sub>2</sub> N <sub>4</sub> O
<b>Molecular Weight:</b>	353.25
<b>Target:</b>	ROCK
<b>Pathway:</b>	Cell Cycle/DNA Damage; Cytoskeleton; Stem Cell/Wnt; TGF-beta/Smad
<b>Storage:</b>	4°C, sealed storage, away from moisture * In solvent : -80°C, 6 months; -20°C, 1 month (sealed storage, away from moisture)



### SOLVENT & SOLUBILITY

<b>In Vitro</b>	DMSO : 100 mg/mL (283.09 mM; Need ultrasonic)					
	H <sub>2</sub> O : 50 mg/mL (141.54 mM; Need ultrasonic)					
	<b>Preparing Stock Solutions</b>	<b>Solvent</b>	<b>Mass</b>	<b>1 mg</b>	<b>5 mg</b>	<b>10 mg</b>
		<b>Concentration</b>				
		<b>1 mM</b>		2.8309 mL	14.1543 mL	28.3086 mL
<b>5 mM</b>			0.5662 mL	2.8309 mL	5.6617 mL	
	<b>10 mM</b>		0.2831 mL	1.4154 mL	2.8309 mL	
Please refer to the solubility information to select the appropriate solvent.						
<b>In Vivo</b>	1. Add each solvent one by one: PBS Solubility: 100 mg/mL (283.09 mM); Clear solution; Need ultrasonic					
	2. Add each solvent one by one: 10% DMSO >> 40% PEG300 >> 5% Tween-80 >> 45% saline Solubility: ≥ 2.5 mg/mL (7.08 mM); Clear solution					
	3. Add each solvent one by one: 10% DMSO >> 90% (20% SBE-β-CD in saline) Solubility: ≥ 2.5 mg/mL (7.08 mM); Clear solution					
	4. Add each solvent one by one: 10% DMSO >> 90% corn oil Solubility: ≥ 2.5 mg/mL (7.08 mM); Clear solution					

### BIOLOGICAL ACTIVITY

<b>Description</b>	Y-33075 dihydrochloride is a selective ROCK inhibitor with an IC <sub>50</sub> of 3.6 nM.		
<b>IC<sub>50</sub> &amp; Target</b>	ROCK 3.6 nM (IC <sub>50</sub> )	PKC 420 nM (IC <sub>50</sub> )	CaMKII 810 nM (IC <sub>50</sub> )
<b>In Vitro</b>	Y-33075 (Y-39983) is a potent ROCK inhibitor, with an IC <sub>50</sub> of 3.6 nM. Y-33075 also inhibits PKC and CaMKII more potently		

than Y-27632, and the IC<sub>50</sub>s of Y-27632 and Y-33075 for PKC are 9.0 μM and 0.42 μM, respectively, whereas the IC<sub>50</sub>s of Y-27632 and Y-33075 for CaMKII are 26 μM and 0.81 μM, respectively. The IC<sub>50</sub>s of Y-27632 and Y-33075 for PKC is 82 and 117 times those for ROCK, respectively, whereas the IC<sub>50</sub>s of Y-27632 and Y-33075 for CaMKII is 236 and 225 times those for ROCK, respectively<sup>[1]</sup>. Y-33075 (Y-39983, 10 μM) extends neurites in the retinal ganglion cells (RGCs) compared with those in RGCs treated without Y-39983<sup>[2]</sup>. Y-33075 (Y-39983, 1 μM) inhibits the contraction of rabbit ciliary artery segments evoked by histamine in Ca<sup>2+</sup>-free solutions. Y-33075 (10 μM) shows no effect on the [Ca<sup>2+</sup>]<sub>i</sub> increase with the high-potassium (high-K) solution<sup>[3]</sup>.

MCE has not independently confirmed the accuracy of these methods. They are for reference only.

#### In Vivo

In rabbits, Y-39983 (≥0.01%) significantly lowers intraocular pressure (IOP) at 2 hours after topical administration. In monkeys, Y-39983 (0.05%)-treated eyes show significant reduction of IOP between 2 and 7 hours after topical administration<sup>[1]</sup>. Y-39983 (100 μM) increases the regenerating axons of retinal ganglion cells (RGCs) in the eyes of the rats<sup>[2]</sup>.

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## PROTOCOL

### Kinase Assay<sup>[1]</sup>

Recombinant ROCK (ROK α/ROCK II), purified protein kinase C (PKC: mixture of α, β, γ isoforms), and recombinant calmodulin-dependent protein kinase II (CaMK II) are used in the assay. ROCK (0.2 U/mL) is incubated with 1 μM [γ-<sup>32</sup>P] ATP and 10 μg/mL histone as substrates in the absence or presence of various concentrations of Y-27632, Y-33075, or staurosporine at room temperature for 20 minutes in 20 mM MOPS (3-(N-morpholino)propanesulfonic acid) buffer (pH 7.2) containing 0.1 mg/mL bovine serum albumin (BSA), 5 mM dithiothreitol [DTT], 10 mM β-glycerophosphate, 50 μM Na<sub>3</sub>VO<sub>4</sub>, and 10 mM MgCl<sub>2</sub> in a total volume of 100 μL. PKC (10 ng/mL) is incubated with 1 μM [γ-<sup>32</sup>P] ATP and 20 μM PKC substrate in the absence or presence of various concentrations of Y-27632, Y-33075, or staurosporine at room temperature for 30 minutes in 20 mM MOPS buffer (pH 7.5) containing 0.1 mg/mL BSA, 10 mM DTT, 10 mM β-glycerophosphate, 50 μM Na<sub>3</sub>VO<sub>4</sub>, 2 mM CaCl<sub>2</sub>, 20 μg/mL phosphatidyl-l-serine, and 10 mM MgCl<sub>2</sub> in a total volume of 100 μL. CaMK II (125 U/mL) is incubated with 1 μM [γ-<sup>32</sup>P] ATP, 10 μM calmodulin, and 20 μM CaMK II substrate, in the absence or presence of various concentrations of Y-27632, Y-33075, or staurosporine at room temperature for 30 minutes in 20 mM MOPS buffer (pH 7.5) containing 0.2 mg/mL BSA, 0.5 mM DTT, 0.1 mM β-glycerophosphate, 50 μM Na<sub>3</sub>VO<sub>4</sub>, 1 mM CaCl<sub>2</sub>, and 5 mM MgCl<sub>2</sub> in a total volume of 100 μL. Incubation is terminated by the addition of 100 μL of 0.7% phosphoric acid. A 160 μL portion of the mixture is transferred to Multiscreen-PH plate. A positively charged phosphocellulose filter absorbs the substrate that binds <sup>32</sup>P. The filter is washed with 300 μL of 0.5% phosphoric acid and then twice with purified water and then dried. The radioactivity of the dried filter is measured with a liquid scintillation counter. Results are presented as 50% inhibitory concentrations and 95% confidence intervals (CIs)<sup>[1]</sup>.

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### Cell Assay<sup>[2]</sup>

In brief, retinal cell suspensions are obtained from dissected retinas of Wistar rats by papain treatment. Retinal ganglion cells (RGCs) are purified by the panning method using anti-rat CD11 antibody for removal of microglia cells and anti-rat Thy-1 antibody for isolation of ganglion cells. The purified RGCs (5000 cells/plate) are seeded into 24-well plates coated by 50 μg/mL of poly-l-lysine and 2 μg/mL of merosin, and are cultured in serum-free neurobasal medium supplemented with 2% B27 supplement, 50 ng/mL BDNF, 50 ng/mL CNTF, 5 μM forskolin, and 1 mM glutamine under a 95% air-5% CO<sub>2</sub> atmosphere at 37°C. After completion of 24-hour cultivation, RGCs are cultured in medium with or without 10 μM Y-33075 as the final concentration for 24 hours and morphologically observed by phase-contrast microscopy. The concentration used is determined based on the effect of Y-33075 on trabecular meshwork contraction in vitro. Since this study is conducted in order to confirm whether Y-33075 has a potential of effect on axonal regeneration of RGCs, the effect is unquantitatively evaluated<sup>[2]</sup>.

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### Animal Administration<sup>[2]</sup>

In brief, SD rats are anesthetized with an intraperitoneal injection of sodium pentobarbital (0.4 mg/kg body weight), and the optic nerve of one eye is transected 4 to 6 mm posterior to the eyeball, taking care to avoid injury to the ophthalmic artery. The anterior branch of the sciatic nerve is excised and sutured autologously to the optic nerve stump with nylon sutures. The other end of the graft is sutured to the temporalis muscle. A small piece (3 mm × 3 mm) of gelatin sponge soaked with 10 μM Y-33075 or saline as a control is implanted in the space behind the optic stump after optic nerve transection in intact

animals. Five  $\mu\text{L}$  of 0.12 mM or 1.2 mM Y-33075 solution or saline is administered into the vitreous body to final concentrations of 10  $\mu\text{M}$  or 100  $\mu\text{M}$ , respectively. The concentrations of Y-33075 used is determined as 10  $\mu\text{M}$  that is effective in the in vitro study on axonal regeneration of RGCs, and also as 100  $\mu\text{M}$  in order to confirm the dose response of Y-33075. Six weeks after surgery, rats is anesthetized with an intraperitoneal injection of sodium pentobarbital (0.4 mg/kg body weight), and 4-Di-10ASP is embedded in the transplanted sciatic nerve to retrogradely label RGCs with axonal regeneration into the sciatic nerve. Three days after dye embedding, rats is euthanized and the eyes is enucleated for preparation of retinal flat-mounts. The posterior eyecup is then separated from the vitreous body and postfixed with 4% paraformaldehyde solution in phosphate buffer for around 1 hour at room temperature. Fluorescence micrographs of the labeled cells is imported using a fluorescence microscope connected to a computer. Labeled cells is counted using image analysis software. As a normal group, the subsequent procedure for retrograde labeling with 4-Di-10ASP is performed without grafting sciatic nerve and administering the test drug. Statistical analysis is performed using logarithmically transformed values due to differences in variance among the groups. The statistical significance of differences between the normal and saline groups and the saline and Y-33075 groups is examined by t-test (onesided) and William's test (one-sided). Findings of  $p < 0.05$  is considered significant<sup>[2]</sup>.

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## CUSTOMER VALIDATION

- Cell. 2018 Jul 26;174(3):636-648.e18.
- Science. 2017 Dec 1;358(6367):eaan4368.
- Nat Commun. 2020 Jan 3;11(1):88.
- Adv Sci (Weinh). 2022 Mar 3;e2104682.
- Stem Cell Rep. 2020 Jan 14;14(1):49-59.

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## REFERENCES

- [1]. Hideki Tokushige, et al. Effects of Topical Administration of Y-39983, a Selective Rho-Associated Protein Kinase Inhibitor, on Ocular Tissues in Rabbits and Monkeys Invest. Ophthalmol. Vis. Sci. July 2007 vol. 48no. 7 3216-3222
- [2]. Tokushige H, et al. Effects of Y-39983, a selective Rho-associated protein kinase inhibitor, on blood flow in optic nerve head in rabbits and axonal regeneration of retinal ganglion cells in rats. Curr Eye Res. 2011 Oct;36(10):964-70.
- [3]. Watabe H, et al. Effects of Rho-associated protein kinase inhibitors Y-27632 and Y-39983 on isolated rabbit ciliary arteries. Jpn J Ophthalmol. 2011 Jul;55(4):411-7. Epub 2011 Jun 11.

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Tel: 609-228-6898

Fax: 609-228-5909

E-mail: [tech@MedChemExpress.com](mailto:tech@MedChemExpress.com)

Address: 1 Deer Park Dr, Suite Q, Monmouth Junction, NJ 08852, USA