CC-401 hydrochloride

| Cat. No.: CAS No.: Molecular Formula: Molecular Weight: Target: Pathway: | HY-13022 1438391-30-0 C ₂₂ H ₂₅ ClN ₆ O 424.93 JNK MAPK/ERK Pathway | |
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| Storage: | 4°C, sealed storage, away from moisture * In solvent : -80°C, 6 months; -20°C, 1 month (sealed storage, away from moisture) | |

SOLVENT & SOLUBILITY

| In Vitro | DMSO : 100 mg/mL (235.33 mM; Need ultrasonic) H ₂ O : 12.5 mg/mL (29.42 mM; Need ultrasonic) | | | | | |
|----------|---|---|--------------------|------------|------------|--|
| | Preparing Stock Solutions | Solvent Mass Concentration | 1 mg | 5 mg | 10 mg | |
| | | 1 mM | 2.3533 mL | 11.7666 mL | 23.5333 mL | |
| | | 5 mM | 0.4707 mL | 2.3533 mL | 4.7067 mL | |
| | | 10 mM | 0.2353 mL | 1.1767 mL | 2.3533 mL | |
| | Please refer to the so | lubility information to select the app | propriate solvent. | | | |
| In Vivo | 1. Add each solvent one by one: PBS Solubility: 14.29 mg/mL (33.63 mM); Clear solution; Need ultrasonic and warming and heat to 60°C | | | | | |
| | 2. Add each solvent one by one: 10% DMSO >> 40% PEG300 >> 5% Tween-80 >> 45% saline Solubility: ≥ 2.5 mg/mL (5.88 mM); Clear solution | | | | | |
| | 3. Add each solvent one by one: 10% DMSO >> 90% (20% SBE-β-CD in saline) Solubility: ≥ 2.5 mg/mL (5.88 mM); Clear solution | | | | | |
| | 4. Add each solvent o Solubility: ≥ 2.5 m | one by one: 10% DMSO >> 90% cor g/mL (5.88 mM); Clear solution | n oil | | | |

| BIOLOGICAL ACTIV | ТТУ |
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| Description | CC-401 hydrochloride is a potent inhibitor of all three forms of JNK with K _i of 25 to 50 nM. |
| IC₅₀ & Target | JNK 25-50 nM (Ki) |
| In Vitro | CC-401 has at least 40-fold selectivity for JNK compared with other related kinases, including p38, extracellular signal- |

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| | regulated kinase (ERK), inhibitor of κB kinase (IKK2), protein kinase C, Lck, zeta-associated protein of 70 kDa (ZAP70). In cell- based assays, 1 to 5 μM CC-401 provides specific JNK inhibition. CC-401, a small molecule that is a specific inhibitor of all three JNK isoforms. CC-401 competitively binds the ATP binding site in JNK, resulting in inhibition of the phosphorylation of the N-terminal activation domain of the transcription factor c-Jun. The specificity of this inhibitor is tested in vitro using osmotic stress of the HK-2 human tubular epithelial cell line. CC-401 inhibits sorbitol-induced phosphorylation of c-Jun in a dosage-dependent manner. However, CC-401 does not prevent sorbitol-induced phosphorylation of JNK, p38, or ERK ^[1] . MCE has not independently confirmed the accuracy of these methods. They are for reference only. |
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| In Vivo | The staining of p-JNK is moderately induced in bevazicumab and Oxaliplatin treatments as compared to control, and in the CC-401-treated samples p-cJun content is significantly lower, consistent with effective JNK inhibition. DNA damage is modestly elevated in combined treatments with CC-401 ^[2] . CC-401 treatment from days 7 to 24 slows the progression of proteinuria, which is significantly reduced compared to the no-treatment and vehicle groups at days 14 and 21. However, there is still an increase in the degree of proteinuria at day 21 in CC-401-treated rats compared to proteinuria at day 5. The vehicle and no-treatment groups developed renal impairment at day 24 as shown by an increase in serum creatinine. This is prevented by CC-401 treatment ^[3] . MCE has not independently confirmed the accuracy of these methods. They are for reference only. |

| Cell Assay ^[1] H | Human HK-2 proximal tubular epithelial cells are cultured in DMEM/F12 media supplemented with 10% FCS, 10 ng/mL EGF, and 10 μg/mL bovine pituitary extract. For Western blot studies, cells are seeded into six-well plates and allowed to adhere avernight, and medium is changed to DMEM/F12 supplemented with only 0.5% FCS for 24 h, by which time cells are |
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| ov cc sc re cu 1 ki M | onfluent. CC-401 is prepared in citric acid (pH 5.5) and added to the confluent cells 1 h before the addition of 300 mM orbitol, and cells are harvested 30 min later using urea-RIPA buffer. Three experiments are performed, each with two eplicates per condition. For ELISA experiments, HK-2 cells are seeded into 24-well plates, allowed to adhere overnight, ultured in DMEM/F12 with 0.5% FCS for 24 h, and then incubated with CC-401 or vehicle for 60 min before stimulation with µM Angiotensin II (AngII). Supernatants are harvested 48 h later and assayed for TGF-β1 content using a commercial ELISA it. Three experiments are performed, each using six replicates per condition ^[1] . |
| Animal M Administration [2][3] To To To <t< td=""><td>Airce^[2] To assess the efficacy of JNK signaling inhibition by CC-401 in anti-angiogenic and Oxaliplatin combination therapy in a nouse xenograft model, adult (8-10 weeks of age) female severe combined immunodeficient mice (C.B.17 SCID) are used. To generate tumors, HT29 cells (1×10⁶ cells) are injected subcutaneously into the left flank of the mice. When the tumors eached approximately 200 mm³, mice are divided into eight groups (eight mice per group) for treatment with Bevacizumab, Oxaliplatin, CC401, and the appropriate combinations of Bevacizumab, Oxaliplatin and CC-401. Mice in the Bevacizumab treatment group receive 5 mg/kg of Bevacizumab by intraperitoneal injection every 3 days for 21 days. The Doxaliplatin treatment group is injected intraperitoneally with 5 mg/kg Oxaliplatin per week for 2 weeks. The CC-401 reatment group is injected intraperitoneally 25 mg/kg for every 3 days. The combination treatment groups receive Bevacizumab (every 3 days, 5 mg/kg), Oxaliplatin (weekly for 2 weeks, 5 mg/kg), and CC-401 (every 3 days, 25 mg/kg). The ontrol group receive saline intraperitoneally. Tumor volume and body weight are measured every 3 days. Tumor volume is alculated. Tumor growth delay is calculated as the difference in the time for control and treated tumors to grow from 200 o 800 mm³. For tumor growth delay calculations, mice are continued to receive treatments till the tumor volume reached 000 m0 m³. For immunohistochemistry mice are sacrificed after treatments on day 9 for tumor processing and staining. tats^[3] iemale WKY rats (180-220 g) are used. Groups of 9 or 10 rats are immunized by subcutaneous injection of 5 mg of sheep IgG n Freund's complete adjuvant followed 5 days later (termed day 0) by a tail vein injection of sheep anti-rat GBM serum. In his study, CC-401 (200 mg/kg/b.i.d. by oral gavage) or vehicle (sodium citrate) treatment is initiated in groups of 9 or 10 rats t 7 days after anti-GBM serum administration and continued twice daily thereafter until animals are killed at da</td></t<> | Airce ^[2] To assess the efficacy of JNK signaling inhibition by CC-401 in anti-angiogenic and Oxaliplatin combination therapy in a nouse xenograft model, adult (8-10 weeks of age) female severe combined immunodeficient mice (C.B.17 SCID) are used. To generate tumors, HT29 cells (1×10 ⁶ cells) are injected subcutaneously into the left flank of the mice. When the tumors eached approximately 200 mm ³ , mice are divided into eight groups (eight mice per group) for treatment with Bevacizumab, Oxaliplatin, CC401, and the appropriate combinations of Bevacizumab, Oxaliplatin and CC-401. Mice in the Bevacizumab treatment group receive 5 mg/kg of Bevacizumab by intraperitoneal injection every 3 days for 21 days. The Doxaliplatin treatment group is injected intraperitoneally with 5 mg/kg Oxaliplatin per week for 2 weeks. The CC-401 reatment group is injected intraperitoneally 25 mg/kg for every 3 days. The combination treatment groups receive Bevacizumab (every 3 days, 5 mg/kg), Oxaliplatin (weekly for 2 weeks, 5 mg/kg), and CC-401 (every 3 days, 25 mg/kg). The ontrol group receive saline intraperitoneally. Tumor volume and body weight are measured every 3 days. Tumor volume is alculated. Tumor growth delay is calculated as the difference in the time for control and treated tumors to grow from 200 o 800 mm ³ . For tumor growth delay calculations, mice are continued to receive treatments till the tumor volume reached 000 m0 m ³ . For immunohistochemistry mice are sacrificed after treatments on day 9 for tumor processing and staining. tats ^[3] iemale WKY rats (180-220 g) are used. Groups of 9 or 10 rats are immunized by subcutaneous injection of 5 mg of sheep IgG n Freund's complete adjuvant followed 5 days later (termed day 0) by a tail vein injection of sheep anti-rat GBM serum. In his study, CC-401 (200 mg/kg/b.i.d. by oral gavage) or vehicle (sodium citrate) treatment is initiated in groups of 9 or 10 rats t 7 days after anti-GBM serum administration and continued twice daily thereafter until animals are killed at da |

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CUSTOMER VALIDATION

- Science. 2017 Dec 1;358(6367):eaan4368.
- Cell Syst. 2018 Apr 25;6(4):424-443.e7.
- J Med Chem. 2023 Mar 6.
- Mol Cancer Res. 2016 Aug;14(8):753-63.
- Harvard Medical School LINCS LIBRARY

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REFERENCES

[1]. Ma FY, et al. A pathogenic role for c-Jun amino-terminal kinase signaling in renal fibrosis and tubular cell apoptosis. J Am Soc Nephrol. 2007 Feb;18(2):472-84.

[2]. Vasilevskaya IA, et al. Inhibition of JNK Sensitizes Hypoxic Colon Cancer Cells to DNA-Damaging Agents. Clin Cancer Res. 2015 Sep 15;21(18):4143-52.

[3]. Ma FY, et al. Blockade of the c-Jun amino terminal kinase prevents crescent formation and halts established anti-GBM glomerulonephritis in the rat. Lab Invest. 2009 Apr;89(4):470-84.

Caution: Product has not been fully validated for medical applications. For research use only.

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